



Draw one aerobic (blue) bottle and one anaerobic (purple) bottle per culture set. See bottom half of this page to determine if green tube is needed.

- Remove caps from culture bottles and disinfect the tops of the culture bottles and green tube with alcohol pad.
- Optimal blood draw volume is 20-22cc. 10cc per culture bottle and 1-2cc for green top tube.
 - To ensure addition of optimal volume, mark with a pen where the ruled lines on the bottle show 10 mL above the liquid media level. Fill to this mark.
 - If unable to obtain optimal blood volume, fill in this order: 1) green top tube if needed (1-2 mL); 2) aerobic (blue) bottle (10 mL); 3) anaerobic (purple) bottle (remainder, if any).
- Write site, date, time, and initials on label. **Do not cover bar code label on bottles!**
- **Do not cover the window with the ruled lines!** (Lab needs to be able to ensure proper blood amounts are in the bottles.)
- Place labels **VERTICALLY** instead of **around** the bottle. (Lab needs to be able to scan the barcode.)
- If using the pneumatic tube system, place green top in separate plastic bag toward the “necks” of the bottles. Place each glass bottle in a separate sealed zip lock bag and then securely double bag the set. **Do not tube more than one set per tube!**

VENIPUNCTURE w/NO CENTRAL LINE (2 CULTURES):

CENTRAL LINE + PERIPHERAL STICK (2 CULTURES) OR 2 CENTRAL LINE DRAWS:

- GREEN top tubes not required.
- Scrub proposed site of venipuncture with Chloraprep™ for 30 seconds using frictional back and forth motion. Allow to air dry naturally for 30 seconds prior to venipuncture.
- Sterile Gloves are NOT necessary. **DO NOT re-palpate the prepared site – this may introduce contamination into the culture!**
- Cultures obtained from the same site must be collected 10 minutes apart; from different sites they may be obtained at the same time.
- ***PRINTING LABELS – 3 labels print out for each culture set whether or not utilizing green tops.***
- Pick 2 labels from the 1st three that print, discard the third label: label purple & blue tops. *Check for matching accession numbers.*
- Pick 2 labels from the 2nd three that print, discard the third label: label purple & blue tops. *Check for matching accession numbers.*

- 2 GREEN top tubes required (1 for Peripheral and 1 for Central).
- Remove current central line cap from line. Scrub the open port with alcohol. Attach **new cap** to lumen using proper sterile technique.
- Disinfect new central line cap with alcohol for 30 seconds. Wait 30 seconds for alcohol to dry.
- Attach 10-20cc luer lock syringe to new cap. Aspirate blood, filling whole syringe. **DO NOT FLUSH line prior to attaching syringe and do not waste any blood!**
- Attach red access transfer device to blood-filled syringe and transfer blood. Allow proper volume of blood to flow passively into each bottle. **Do not force blood into bottles!**
- **Wait at least 10 minutes before obtaining second culture set from the same line.** Make sure to print new stickers at timing of second draw.